

Quantitative Analysis of Interleukin-6 in Vitreous from Patients with Proliferative Vitreoretinal Diseases

Shingo Kojima, Takahiko Yamada and Makoto Tamai

Department of Ophthalmology, Tohoku University School of Medicine, Sendai, Japan

Purpose: To explore immunological mechanisms in the pathogenesis of proliferative vitreoretinal diseases, we measured the concentration of interleukin-6 in the vitreous body and serum from patients with proliferative diabetic retinopathy (PDR), proliferative vitreoretinopathy (PVR), and premacular fibrosis. To evaluate immunological etiology, interleukin-6 levels in each disease were compared with disease severity.

Methods: Clinical samples were obtained at the beginning of pars plana vitrectomy from 30 eyes of 26 patients with PDR, 12 eyes of 12 patients with PVR, and 10 eyes of 10 patients with premacular fibrosis. Interleukin-6 was quantitated with an enzyme-linked immunosorbent assay.

Results: The levels of detectable interleukin-6 in the vitreous specimens ranged from 22.8 to 666.4 pg/mL in the PDR patients and from 28.2 to 416.3 pg/mL in the PVR patients. No interleukin-6 was detected in the vitreous specimens from patients with premacular fibrosis or in any serum samples from patients. Interleukin-6 levels of vitreous specimens from PDR patients were higher than those from PVR patients (P < .02, Mann–Whitney U-test). There was no correlation between clinical severity and interleukin-6 levels in vitreous specimens from either PDR or PVR patients.

Conclusion: Our results indicated that cell-mediated immunity is involved in the pathogenesis of proliferative vitreoretinal diseases. **Jpn J Ophthalmol 2001;45:40–45** © 2001 Japanese Ophthalmological Society

Key Words: Interleukin-6, vitreous, vitreoretinal disease.

Introduction

Preretinal membranes in patients with proliferative vitreoretinal diseases, such as proliferative diabetic retinopathy (PDR) and proliferative vitreoretinopathy (PVR), are thought to be caused by migration and proliferation of various types of cells, including pigment epithelial cells, fibroblasts, glial cells, vascular endothelial cells, and macrophages.¹⁻⁶ It has been reported that these phenomena are modified by various cytokines and growth factors⁷⁻¹⁰ that are released from both normal and damaged retina. When retinal ischemia or retinal detachment occurs, these damaged areas may release cytokines. Many studies have been performed to resolve the pathogenesis of proliferative retinal diseases. Autoimmune mechanisms have been thought to play a role in the pathogenesis of proliferative retinal diseases.^{11–13} Rahi and Addison¹⁴ have suggested a relationship between immunological mechanisms and the development of microangiopathy. Baudouin et al^{15–21} found ocular deposits of immunogloblin, complement, and human leukocyte antigen in patients with PDR and PVR in a series of studies using an immunohistochemical method.

Interleukin-6 is known to be a potent cytokine in cell-mediated immunity that plays an important role in the regulation of immune response, acute phase reactions, and hematopoiesis. Abnormal production of interleukin-6 has been implicated in many kinds of diseases, including autoimmune diseases, chronic inflammation, and lymphoid malignancies. To explore immunological mechanisms in the pathogene-

Received: October 25, 1996

Correspondence and reprint requests to: Shingo KOJIMA, MD, Department of Ophthalmology, Tohoku University School of Medicine, 1-1 Seiryo-machi, Aoba-ku, Sendai-shi, Miyagi-ken 980-8574, Japan

sis of proliferative vitreoretinal diseases, we measured the concentration of interleukin-6 in the vitreous body and serum from patients with PDR, PVR, and premacular fibrosis and compared it with the severity of each disease.

Materials and Methods

Patients

Clinical samples were obtained at the beginning of pars plana vitrectomy from 30 eyes of 26 patients with PDR (Table 1), 12 eyes of 12 patients with PVR (Table 2), and 10 eyes of 10 patients with premacular fibrosis (Table 3). The PDR patients included 17 men and 9 women, ranging in age from 28 to 67 years (average: 52 years). The duration of diabetes in these patients ranged from 2 to 30 years. Of 26 PDR patients, 25 patients had non-insulin-dependent diabetes (type II). Only 1 patient was insulin-dependent

 Table 1.
 Interleukin-6 Levels in Vitreous from Proliferative

 Diabetic Retinopathy (PDR) Patients and Clinical Severity

Patient		Age	Clinical Severity	IL- 6^{\dagger}	HbA1c [‡]
No.	Sex	(y)	(Grade)*	(pg/mL)	(mg/dL)
1	М	50	1	666.4	6.7
2	Μ	39	1	141.9	NA
3	F	62	2	223.4	5.3
			2	97.9	NA
4	Μ	57	2	167.3	NA
5	F	51	1	136.9	7.8
6	F	45	1	229.9	10.5
7	F	46	1	50.5	5.8
8	F	43	2	ND	7.1
9	F	49	2	73.1	7.1
			2	77.1	NA
10	Μ	56	2	ND	7.7
11	Μ	67	1	116.7	6.9
12	Μ	43	2	58.5	7.1
13	F	56	3	51.1	7.4
			3	43.2	NA
14	Μ	61	1	81.9	8.1
15	Μ	52	1	207.2	5.6
16	Μ	55	1	103.7	6.7
17	Μ	57	1	96.6	7.1
18	Μ	52	1	134.7	NA
19	Μ	60	1	53.4	6.7
20	Μ	41	0	45.4	9.8
21	Μ	51	2	24.9	7.1
22	Μ	54	2	171.9	9.2
23	Μ	28	3	208.6	6.1
24	F	59	3	171.6	11.8
25	F	65	1	22.8	6.7
26	Μ	50	2	73.5	NA
			2	73.5	NA

^{*}Clinical severity according to method of Abu el Asrar et al. [†]ND: Not detectable.

Patient		Age	Clinical	IL-6
No.	Sex	(ÿ)	Severity*	$(pg/mL)^{\dagger}$
1	М	57	D1	111.9
2	F	64	C2	ND
3	Μ	19	D1	65.8
4	Μ	44	D1	92.7
5	Μ	24	C1	ND
6	F	63	В	ND
7	Μ	18	D1	ND
8	F	62	D1	28.2
9	Μ	23	D3	86.8
10	М	48	C2	416.3
11	М	6	C2	ND
12	Μ	58	В	36.7

Table 2. Interleukin-6 Levels in Vitreous from Proliferative

Vitreoretinopathy (PVR) Patients

*Clinical severity according to Retina Society Terminology Committee.

[†]ND: Not detectable.

(type I). The clinical severity of PDR was classified from 0 to 4, according to the grading of Abu el Asrar et al.²² Grading was based on the presence and extent of active fibrovascular tissue, vitreous hemorrhage, iris neovascularization, and hyphema. Grade 0 indicated inactive residual fibrous tissue from proliferative diabetic retinopathy; grade 1+, vitreous hemorrhage without active neovascularization; grade 2+, vitreous hemorrhage with actively growing fibrovascular tissue; grade 3+, iris neovascularization in addition to the findings in grade 2+; and grade 4+, hyphema in addition to the findings in grade 3+. The PVR patients included 9 men and 3 women, ranging in age from 6 to 64 years (average: 40 years). The clinical severity of PVR was classified by the grading reported by the Retina Society Terminology Committee.²² The premacular fibrosis patients included 5 men and 5 women, ranging in age from 13

Table 3. Interleukin-6 Levels in Vitreous from PremacularFibrosis (PMF) Patients

Patient No.	Sex	Age (y)	PMF Diagnosis	IL-6 (pg/mL)*
1	F	72	Idiopathic	ND
2	F	59	Idiopathic	ND
3	М	42	Idiopathic	ND
4	F	50	Idiopathic	ND
5	F	76	Idiopathic	ND
6	Μ	58	Secondary	ND
7	Μ	60	Secondary	ND
8	Μ	65	Secondary	ND
9	Μ	13	Secondary	ND
10	F	50	Secondary	ND

*ND: Not detectable.

[‡]NA: Not available.

to 75 years (average: 60 years). Five of these patients had idiopathic premacular fibrosis, while the conditions of the others were secondary to cryoretinopexy and scleral encircling procedures. Patients who had previous vitrectomy or inflammatory diseases were excluded from this study. Informed consent was obtained from each patient before surgery.

Collection of Vitreous and Serum Specimens

Collection of vitreous and serum specimens was performed at the beginning of pars plana vitrectomy, as described previously.²³ In brief, vitreous specimens were collected in a glass bottle that had an airtight rubber cap and arms. The bottle was connected via the arms to the spliced suction tube of a vitreous cutter (Alcon, Ft. Worth, TX, USA). The infusion line was first filled with air, and then core vitrectomy was carefully done. Air was infused at the pressure level of 40 to 55 mm Hg. Thus, undiluted vitreous fluid was collected in the bottle. After a sufficient amount of vitreous was obtained safely, the vitreous cutter was removed from the eyeball, and the air in the vitreous cavity was replaced by irrigating solution (BSS; Alcon). The collecting bottle was disconnected from the suction line and was placed immediately in ice-cold water. After the vitreous specimen was obtained, venous blood was sampled and serum was prepared by centrifugation. The vitreous specimen and serum were aliquoted in micro test tubes and frozen at -70° C until the enzyme assay.

Quantification of Interleukin-6

Interleukin-6 was quantitated with an enzymelinked immunosorbent assay kit (Serotec; Blackthorn, UK). Duplicate aliquots of 100 μ L of undiluted vitreous and serum were tested. The standard curve was obtained with recombinant human interleukin-6, ranging from 12.5 to 800 pg/mL. The detection limit was 16 pg/mL.

Statistical Analysis

Data were analyzed by using the Mann–Whitney U-test. A P-value <.05 was considered significant. Values were expressed as mean \pm SEM.

Results

Interleukin-6 levels could be measured in 28 of 30 vitreous specimens (93%) from patients with PDR and in 7 of 12 specimens (58%) from patients with PVR (Tables 1 and 2). In the vitreous specimens from patients with premacular fibrosis, no interleu-

kin-6 was detected (Figure 1). The levels of detectable interleukin-6 in the vitreous specimens ranged from 22.8 to 666.4 pg/mL (136.1 \pm 26.1 pg/mL; mean \pm SEM) in the PDR patients and from 28.2 to 416.3 pg/ mL (69.9 \pm 33.7 pg/mL; mean \pm SEM) in the PVR patients. In none of the serum samples was interleukin-6 detected. Interleukin-6 levels of vitreous specimens from PDR patients were significantly higher than those from PVR patients (P < .02, Mann–Whitney U-test) (Figure 2). There was no correlation between interleukin-6 levels in the vitreous specimen from PDR patients and the grade of clinical severity (Figure 3). There also was no correlation in PVR patients between interleukin-6 levels in vitreous specimens and clinical severity (Figure 4).

Discussion

Preretinal membranes of PDR patients contain newly formed vessels and extracellular matrix that were affected by liberated angiogenic factors produced by the ischemic retina, so-called nonperfusion areas. The characteristics of these angiogenic factors involved in retinal angiogenesis have not yet been clarified.

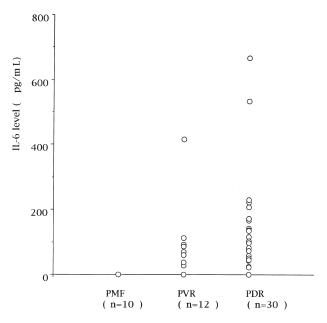


Figure 1. Interleukin-6 levels in vitreous from patients with proliferative diabetic retinopathy (PDR), proliferative vitreoretinopathy (PVR), and premacular fibrosis (PMF). No interleukin-6 levels could be detected in any vitreous samples from premacular fibrosis patients, nor in 5/12 samples from PVR patients, or in 2/30 samples from PDR patients.

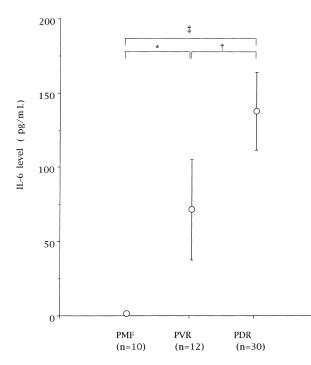


Figure 2. Interleukin-6 levels in vitreous from patients with proliferative diabetic retinopathy (PDR), proliferative vitreoretinopathy (PVR), and premacular fibrosis (PMF). Each point represents mean \pm SE. Interleukin-6 levels in vitreous from PDR patients were higher than those from PVR patients. **P* < .05, [†]*P* < .02, [‡]*P* < .0001 (Mann–Whitney *U*-test)

The main complication of rhegmatogenous retinal detachment is PVR. This complication results from migration and proliferation of various types of cells, including those derived from pigment epithelium, glial cells, fibroblast-like cells, and macrophages.¹⁻⁶ Although the pathogenesis of these two diseases are different in many aspects, both PDR and PVR have similar cellular processes; for example, cell activation, migration, proliferation, and synthesis of extracellular matrix. All these phenomena are recently reported to be induced by various cytokines and growth factors,⁷⁻¹⁰ but the roles of these mediators in these two diseases remain speculative.

Interleukin-6 is a multifunctional cytokine; its functions include induction of the final differentiation of B cells to antibody-producing cells; induction of T cells to killer T cell activation; interleukin-2 receptor expression; proliferation of IL-6 itself; induction of acute phase protein production; and suppression of albumin production. This cytokine also is known to be a mediator in cell-mediated immunity, which increases in the synovial fluid or in the serum of patients with autoimmune disease, such as rheu-

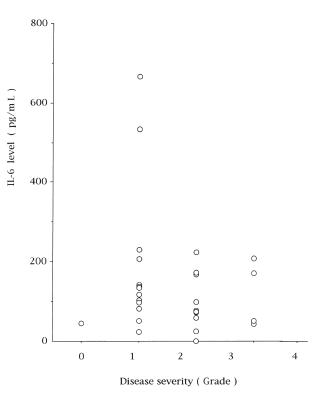
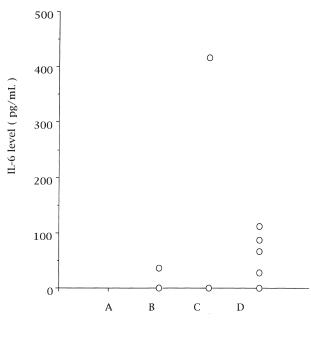


Figure 3. Interleukin-6 levels in vitreous from patients with proliferative diabetic retinopathy (PDR). There was no correlation between interleukin-6 levels in 30 vitreous samples from PDR patients and grade of clinical severity.

matoid arthritis.^{24–26} Interleukin-6 levels correlate with autoimmune disease activity.²⁶

In recent studies,^{11–13} cell-mediated immunity has played an important role in the pathogenesis of proliferative retinopathy. Biopsies of the pars plana in PDR and PVR patients have demonstrated the presence of immunogloblin, deposits of complement, and human leukocyte antigen expression at the level of the pigmented epithelium. These deposits and expression have been detected in the preretinal membranes in the connective stroma and within the vascular walls.¹⁵⁻²⁰ Under normal conditions, these antigens are restricted to immunocompetent cells and play regulatory roles in the interactions among the various cells of the immune system. Deviant expression of class II antigen has been found on nonlymphoid cells in various autoimmune diseases.^{27,28} From the present study, it has been speculated that ischemic or detached retina in proliferative vitreoretinal disease has autoimmunity that causes an excessive wound-healing process. This excessive process may be involved in the pathogenesis of proliferative vitreoretinal diseases.



Disease severity (Grade)

Figure 4. Interleukin-6 levels in vitreous from patients with proliferative vitreoretinopathy (PVR). There was no correlation between interleukin-6 levels in 12 vitreous samples from PVR patients and grade of clinical severity.

Cell-mediated immunity is thought to be involved in the pathogenesis of acute retinal necrosis (ARN). We measured the concentration of interleukin-6 in the vitreous from 3 eyes of 3 patients with ARN. The levels of interleukin-6 were high, ranging from 602.4 to 1142.8 pg/mL (864.2 \pm 156.2 pg/mL; mean \pm SEM). We think that these levels may reflect intense cell-mediated immunity in inflammatory disease.

Interleukin-6 levels in the vitreous of patients with proliferative retinopathy have been reported by Abu el Asrar et al.²⁴ They reported that interleukin-6 levels in the vitreous from PDR patients increased, and they found correlations between the level of this cytokine in the vitreous and the grade of clinical severity. Kauffmann et al²⁹ reported that interleukin-6 levels correlated with clinical severity in PVR patients. In this study, interleukin-6 levels in the vitreous from both PDR and PVR patients increased. The PDR patients had higher interleukin-6 levels in the vitreous than did PVR patients.

Our results were somewhat different from those of Abu el Asrar et al²¹ and Kauffmann et al.²⁹ We collected not only the soluble fraction of the vitreous but also the gel fraction by using the vitreous cutter. It is, however, unclear what difference the sampling method may have on interleukin-6 levels in the vitreous. But it is a fact that interleukin-6 levels were increased in PDR and PVR patients using our method. The results of our current study indicate that cellmediated immunity may be involved in the pathogenesis of proliferative vitreoretinal diseases.

Our results showed that ischemic retinal damage in PDR may be much more severe than detached retinal damage in PVR from the viewpoint of stimulation of interleukin-6 production. There was no correlation between interleukin-6 levels in the vitreous and clinical ocular severity in both PDR and PVR patients. We think the present grading of clinical severity for PDR and PVR does not reflect the intensity of cell-mediated immunity. A new classification for these diseases may be necessary to evaluate the activation level of intraocular immunity.

References

- 1. Van HD, Aaberg TM, Machemer R, Fenzl R. Glial cell proliferation in human retinal detachment with massive periretinal proliferation. Am J Ophthalmol 1977;84:383–93.
- Machemer R, Van HD, Aaberg TM. Pigment epithelial proliferation in human retinal detachment with massive periretinal proliferation. Am J Ophthalmol 1978;85:181–91.
- Newsome DA, Rodrigues MM, Machemer R. Human massive periretinal proliferation. In vitro characteristics of cellular components. Arch Ophthalmol 1981;99:873–80.
- Hiscott PS, Grierson I, McLeod D. Retinal pigment epithelial cells in epiretinal membranes: an immunohistochemical study. Br J Ophthalmol 1984;68:708–15.
- Ryan SJ. The pathophysiology of proliferative vitreoretinopathy in its management. Am J Ophthalmol 1985;100:188–93.
- Weller M, Heimann K, Wiedemann P. Demonstration of mononuclear phagocytes in a human epiretinal membrane using a monoclonal anti-human macrophage antibody. Graefes Arch Clin Exp Ophthalmol 1988;226:252–4.
- Kirchhof B, Sorgente N. Pathogenesis of proliferative vitreoretinopathy. Modulation of retinal pigment epithelial cell functions by vitreous and macrophages. Dev Ophthalmol 1989;16:1–53.
- Wiedemann P, Weller M, Heimann K. Proliferative vitreoretinopathy: new discoveries in pathophysiology and therapy. Klin Monatsbl Augenheilkd 1990;197:355–61 (in German).
- Fredj-Reygrobellet D, Baudouin C, Negre F, Caruelle JP, Gastaud P, Lapalus P. Acidic FGF and other growth factors in preretinal membranes from patients with diabetic retinopathy and proliferative vitreoretinopathy. Ophthalmic Res 1991;23:154–61.
- 10. Wiedemann P. Growth factors in retinal diseases: proliferative vitreoretinopathy, proliferative diabetic retinopathy, and retinal degeneration. Surv Ophthalmol 1992;36:373–84.
- 11. Henley WL, Okas S, Leopold IH. Leukocyte migration inhibition by chroid and retina in retinal detachment. Opthalmic Res 1975;7:129–32.
- Brinkman CJ, Broekhuyse RM. Cell-mediated immunity after retinal detachment as determined by lymphocyte stimulation. Am J Ophthalmol 1978;86:260–5.
- Remy C. Autoimmunity against the retina in idiopathic retinal detachment. Study of 50 cases. J Fr Ophtalmol 1981;4:213–7 (in French).

- Rahi AH, Addison DJ. Autoimmunity and the outer retina. Trans Ophthalmol Soc UK 1983;103:428–437.
- Baudouin C, Fredj-Reygrobellet D, Lapalus P, Gastaud P. Immunohistopathologic findings in proliferative diabetic retinopathy. Am J Ophthalmol 1988;105:383–8.
- Baudouin C, Fredj-Reygrobellet D, Baudouin F, Lapalus P, Gastaud P. Immunohistologic study of proliferative vitreoretinopathy. Am J Ophthalmol 1989;108:387–94.
- Baudouin C, Fredj-Reygrobellet D, Jambou D, Righini M, Gastaud P. Immunopathologic study of retinal detachment with vitreo-retinal proliferation. Ophtalmologie 1989;3:22–5 (in French).
- Baudouin C, Gordon WC, Fredj-Reygrobellet D, et al. Class II antigen expression in diabetic preretinal membranes. Am J Ophthalmol 1990;109:70–4.
- Baudouin C, Gastaud P, Gordon B, Lapalus P, et al. Immunohistologic study of the epiretinal membranes in proliferative diabetic retinopathy and retinal detachment with vitreo- retinal proliferation. Ophtalmologie 1990;4:53–5 (in French).
- Baudouin C, Fredj-Reygrobellet D, Brignole F, Lapalus P, Gastaud P. MHC class II antigen expression by ocular cells in proliferative diabetic retinopathy. Fundam Clin Pharmacol 1993;7:523–30.
- Abu el Asrar, MA, Maimone D, Morse PH, Gregory S, Reder AT. Cytokines in the vitreous of patients with proliferative diabetic retinopathy. Am J Ophthalmol 1992;114:731–6.

- 22. Retina Society Terminology Committee. The classification of retinal detachment with proliferative vitreoretinopathy. Oph-thalmology 1983;90:121–5.
- Tamai M, Nakazawa M. A collection system to obtain vitreous humor in clinical cases (letter). Arch Ophthalmol 1991;109:465–6.
- 24. Hirano T, Matsuda T, Turner M, et al. Excessive production of interleukin 6/B cell stimulatory factor-2 in rheumatoid arthritis. Eur J Immunol 1988;18:1797–801.
- 25. Bhardwaj N, Santhanam U, Lau LL, et al. IL-6/IFN-beta 2 in synovial effusions of patients with rheumatoid arthritis and other arthritides. Identification of several isoforms and studies of cellular sources. J Immunol 1989;143:2153–9.
- Waage A, Kaufmann C, Espevik T, Husby G. Interleukin-6 in synovial fluid from patients with arthritis. Clin Immunol Immunopathol 1989;50:394–8.
- Bottazzo GF, Pujol BR, Hanafusa T, Feldmann M. Role of aberrant HLA-DR expression and antigen presentation in induction of endocrine autoimmunity. Lancet 1983;2:1115–9.
- Foulis AK, Farquharson MA. Aberrant expression of HLA-DR antigens by insulin-containing beta-cells in recent-onset type I diabetes mellitus. Diabetes 1986;35:1215–24.
- Kauffmann DJ, van Meurs JC, Mertens DA, Peperkamp E, Master C, Gerritsen ME. Cytokines in vitreous humor: interleukin-6 is elevated in proliferative vitreoretinopathy. Invest Ophthalmol Vis Sci 1994;35:900–6.